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Dielectric Modulated Charge Plasma-Based Label Free Detection of Biomolecules in Dual Metal DG TFET Biosensors

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ABSTRACT In this article, we present dielectric modulation-based label free detection of biomolecules at a cavity under gate metal located near the tunneling junction of charge plasma based dual metal-double gate tunnel FET (CP-DM DGTFET). The biomolecules that settle down in the cavity act as dielectric above the tunneling junction and cause drain current to increase/ decrease. To eliminate the short channel impact and to enhance I_{ON}/I_{OFF} ratio without compromising any other device features, dual metal gate architecture with laterally split dielectric is used. The biosensor is capable of recognizing neutral, positive, and negative charges with the highest drain current sensitivity of 2.9×10^9 Ccm⁻² and highest I_{ON}/I_{OFF} ratio of 9.02×10^{11} for biomolecule gelatin with dielectric constant k = 12. This work explores the effectiveness of charge-plasma based DM DGTFET for detection of biomolecules such as gelatin (k = 12), keratin (k = 8), bacteriophage T7 (k = 6.3), and APTES (k = 3.57) with varied charge distribution at various biasing. The drain current sensitivity, subthreshold slope, drain current, ON/OFF current ratio are computed for various biomolecules and their variations with respect to cavity length, biomolecule charge, etc. are used to detect unknown biomolecules. Finally, RF analysis such as gain bandwidth product, cut-off frequency, transit time, and linearity/selectivity of biosensors are incorporated. The device shows better linearity and less distortion for high –k biomolecules. The results of our studies are reproducible for repeated computed analysis.

INDEX TERMS Biosensor, charge plasma, dielectric modulation, sensitivity, split dielectric.

I. INTRODUCTION

MOSFET with nanogap has been a promising device to detect label free biomolecules. The reason behind its popularity lies in its improved detecting capacity, minimal power usage, low cost, and CMOS compatibility. Despite these benefits,

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FET-based biosensors have inevitable issues such as poor subthreshold swing (SS > 60 mV/dec), restricted sensitivity, elevated power dissipation caused by thermionic electron emission and short channel effects (SCEs). The technique popularly used these days is dielectric modulation [1], [2], where alterations in the gate-oxide dielectric constant (k) result in changes in the sensitivity of the biosensor. To eradicate these, researchers turned their attention to TFET that

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focuses on tunneling process of carriers from band to band, controlled charge transport mechanism and a steep subthreshold swing. Furthermore, label-free detection is a key feature of TFET biosensors, allowing direct observation of biomolecular interactions without requiring markers. This technique complements the device's precision charge transport management and sharp subthreshold swing by streamlining the detection procedure and increasing sensitivity. Detection and analysis of biomolecules are important for diseases diagnostic, food safety etc. Any change in concentration of some specific biomolecules may lead to malfunction of living cells. Accurate measurement of specific biomolecule concentration in living cells are very crucial to ensure optimum functioning of living organisms.

Tunnel FET based biosensors are one of the most fascinating platforms demonstrating promising potential for an electrical detection of label-free and charged biomolecules by their intrinsic charges. We have illustrated in this article how TFET based biosensors open up new possibilities for fast and on-site medical diagnostics for healthcare solutions. The analysis focuses on methodologies aiming to improving sensitivity and assessing shifts in simulated parameters such as ON current (ION), OFF current (IOFF), ON-OFF current ratio (I_{ON}/I_{OFF}), Subthreshold Swing (SS), threshold voltage (V_{th}), sensitivity, and selectivity. Doping-less chargeplasma-based FET [3], [4], [5], [6] is the most popular among TFET based sensors and it also eased the process of fabrication by creating drain (N⁺ region) and source (P⁺ region) with appropriate work functions with respective drain and source metal contacts.

The benefits of dual metal double gate electrodes, specifically M1 (such as hafnium that forms the tunneling gate with lower work function ϕ_t to trigger the early tunneling phenomena at source-channel side) and M2 (such as aluminium that forms the auxiliary gate with higher work function $\phi_a > \phi_t$ to swipe the charges towards the drain), have also been integrated to enhance the device performance [7]. In TFETs, using a metal with a lower work function can help decrease the turn-on voltage. This happens because a smaller work function enhances the bending of the energy bands at the source-channel interface, which promotes more efficient electron tunneling and improves performance at lower voltages. While a metal gate having higher work function at the drain-channel junction reduces the OFF current by increasing the energy barrier, which limits electron leakage when the device is in the OFF state. Also, this leads to a decrease in ambipolarity behaviour, hindrance of short channel effects, and reduced drain-induced barrier lowering (DIBL). Girish et al [8] proposed dual metal charge plasma TFET (DM CPTFET) biosensor with a hetero gate structure. For various dielectrics, they conducted sensitivity analysis. Dibyendu et al [9] found superior sensitivity performances of hetero dielectric as compared to single dielectric. Utilizing low-k dielectric closer to the drain and high-k dielectric close to the source [10], [11] as well as better gate modulation at the channel-source intersecting tunnel junction would help reduce ambipolar conduction. To enhance the ON and OFF state electrostatics, high-k spacers have been explored for optimization of digital and analog performance [12], [13]. Recent research show that subthreshold swing (SS) is more improved where ferroelectric material is used as gate dielectric [14], [15], [16]. Some architectures employ diagonal tunneling, providing superior junction electrostatics and gate control [17] while some architectures implement both vertical and lateral tunneling [18]. Recently, using chalcogenide thin films in graphene-based Surface Plasmon Resonance (SPR) sensors has shown enhanced sensitivity and precision in detecting biomolecular interactions. Different ranges of doping techniques in junctionless devices [19] and placement of biomolecules with respect to tunneling junction [20] have proved significant impact for various detectability parameters of biomolecules. It has become evident that the sensitivity of biosensors depends on cavity length [21], [22], [23], drain doping, gate underlap/ overlap [24], multigate configuration [25], additional/extended cavity region [26], [27], [28], additional core gate [29], different gate shapes [30], [31] and biomolecule charge types [32]. The heterojunction TFET architecture [33], [34], [35] shows the suppression of ambipolar conduction and outperform in analog/DC operations. The effect of gate electrode length and metal electrode work function variation has been analyzed in for improving bandto-band tunneling efficiency [11]. In the context of detecting biomolecules, the studies of some parameters such as limit of detection (LOD), limit of quantification (LOQ), linearity, sensitivity, selectivity and reproducibility are very crucial. LOD refers to the minimum concentration of a biomolecule that can be reliably distinguished and are calculated using standard deviation of the response curve and the slope of the calibration curve. LOQ is the same as the LOD if predefined objectives are met. For the reliable and fast detection, it is expected that the characteristic parameters should show linear variation with respect to the concentration of analytes such as biomolecules. Linearity limit defines the operating range between the LOQ and the point where the plot goes nonlinear. The correctness of the observation is decided by the term reproducibility which shows the ability of the biosensor to produce identical responses for a repeated experimental setup. This parameter of the sensor is characterized by the accuracy and precision of the associated electronics circuits.

In this proposed structure, a nanocavity has been created under the M1 gate electrode to detect biomolecules like proteins, amino acids, etc. DNA molecules with polarization and the ability of dipole movements act as both negative and positive charge can also be detected by using the sensor in a high end bioengineering laboratory. This structure offers improved sensitivity for neutral and charged biomolecules. The variation in electrical characteristics, namely, drain current $I_{\rm d}$, $I_{\rm ON}/I_{\rm OFF}$ ratio, sensitivity and device efficiency are captured to identify biomolecules with different dielectric values and charge density. To avoid capacitive coupling of gate over the



tunneling junction, an opening close to gate is used that acts as inlet to the biomolecules towards the cavity. The device's performance is evaluated using the Silvaco ALTAS device simulator.

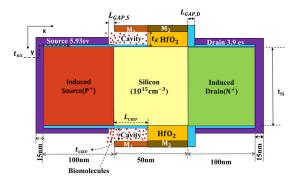


FIGURE 1. Cross-section of dielectric modulated charge plasma based DM DGTFET.

II. DESIGN OF SENSOR

Fig. 1 presents the cross-sectional view of the dielectric modulated charge plasma-based dual metal double gate TFET. The parameters of the device include a single Silicon layer (t_{Si}) of 10 nm thick with intrinsic carrier concentration $n_i=10^{15}~{\rm cm}^{-3}$, channel length $L_T=50$ nm, with different metal work functions above the channel. In this charge plasma-based structure, the drain and source are developed by surrounding the regions with the optimum work function of 3.9 eV and 5.93 eV, respectively [7], [37]. Considering the compliance with Debye's length for optimal performance that avoids quantum effects, a 10 nm silicon body thickness (t_{Si}) is used.

In Charge Plasma based DM DGTFET, two different dielectrics are used as the gate dielectric. The tentative process flow of the given biosensor with a nanogap cavity using standard IC fabrication techniques is illustrated in Figure 2 and elaborated as follows: (1) Initially, an intrinsic (100) silicon wafer with a substrate thickness of 10 nm is considered. (2) A layer of HfO2 is deposited at the gate region cavity using atomic layer chemical vapor deposition with high K and then patterned using chemical mechanical planarization (CMP). (3) An atomic layer of SiO₂ (0.5 nm) is formed on the silicon surface through precise oxidation exposure. (4) Dummy oxidation is performed to shield the cavity and spacer regions at the source end from metal deposition. (5) Certain sections of the insulator are eliminated away using dry etching. (6) Metallization is carried out using a lower-function metal to provide the source electrode, drain electrode, and gate region. (7) Additional metals from the spacer are etched to provide insulation from gates. (8) Selective regions (source electrode, auxiliary gate metal) are exposed to an annealing process with the desired metal gas to achieve different work functions. (9) The sacrificial layer deposited at the cavity and inlet regions is etched away to create the cavity region. (10) Two identical wafers are joined together using the wafer bonding technique to provide dual cavity regions. (11) The bonded wafer forms the desired biosensor device.

Table 1 depicts the permittivity of various biomolecules under study along with the source of references. The dual metal gate allows higher electrostatic capacity to regulate the channel by enabling alignment of the energy bands. Based on the IRDS 2022 roadmap, 0.5 nm SiO₂ equivalent oxide thickness (EOT) is employed below the cavity under tunneling gate and a high-k dielectric (HfO₂) is employed under the auxiliary gate. The region below the tunnel gate metal and above the 0.5 nm SiO₂ layer forms the cavity that acts as trapping site for targeted biomolecules [38]. Due to the differences in physical thickness of laterally split dielectrics, a cavity structure is created below the tunneling gate. The cavity is surrounded by source electrode (work function 5.93 eV) on its left, high-k dielectric (HfO₂) on the right, tunnel gate metal M1 on the top and 0.5 nm SiO₂ layer on the bottom. As soon as biomolecules build up within the cavity, an overall dielectric constant at channel-source intersecting region changes which results in energy band bending and ensures onset of the tunneling phenomena.

TABLE 1. Biomolecules with respective dielectric constants.

Biomolecules	DIELECTRIC CONSTANT (K)	
APTES	3.57[38]	
Bacteriophage T7	6.3[39]	
Keratin	8[40]	
Gelatin	12[41]	

The cavity capacitance C_{cav} is given by

$$Ccav = \frac{\varepsilon_{SiO_2} * k}{k * t_{ox} + \varepsilon_{SiO_2} t_{cav}}$$
(1)

where ε_{SiO2} and k are the permittivity of SiO₂ layer, and the biomolecule, respectively, tox being the thickness of SiO₂ layer. Here fully filled cavity is considered for investigation with the cavity dimensions (length, L_{cav} and thickness, t_{cav}). The cavity under study has the dimensions of 20 nm × 2.7 nm. The influence of cavity dimensions on the ability to sense biomolecules is being investigated here. The calculation is repeated for other dimensions of cavity 12 nm ×2.5 nm. Here fully filled cavity is considered for investigation with the cavity dimensions (length, L_{cav} and thickness, t_{cav}). The cavity under study has the dimensions of 20 nm × 2.7 nm. The influence of cavity dimensions on the ability to sense biomolecules is being investigated here. The calculation is repeated for other dimensions of cavity 12 nm ×2.5 nm. The immobilization of biomolecules is facilitated by a 3 nm gap (L_{GAP,S}) between gate and source electrodes. The spacer width between the gate and drain electrode (L_{GAP,D}) has been optimized [43], [44] and is inserted with 15 nm thick layer of SiO2 layer. The tunneling gate is placed over the cavity, as pointed out in Fig. 2. Table 2 gives the device specification of the given structure. Here fully filled cavity is considered for investigation with the



cavity dimensions (length, L_{cav} and thickness, t_{cav}). The 2D simulator named Silvaco ATLAS is used to aid the simulation of CP DM DGTFET [45].

TABLE 2. Device parameters specification.

Specification	Symbol	Parameter values
Length of total gate region, nm	L_{T}	50
Length of cavity region, nm	L_{cav}	20
Thickness of cavity region, nm	t_{cav}	2.7
Tunnel gate work-function, eV	$oldsymbol{\phi}_t$	3.9
Auxiliary gate work-function, eV	ϕ_a	4.5
Spacer thickness, nm	L _{GAP.S.} L _{GAP.D}	3, 15
SiO ₂ thickness, Å	t _{ox}	5
High dielectric thickness, EOT	t_{k}	0.5

The distinct models considered are the bandgap narrowing, Auger models, non-local band-to-band tunneling model, Fermi–Dirac statistics Lombardi (CVT) model, band-to-band quantum tunneling, Shockley–Read–Hall (for recombination) and field-dependent mobility model. To study the way, it performs in the OFF-state, the simulation also incorporates the trap-assisted tunneling model. Moreover, to evaluate the numerical tunneling probability Wentzel-Kramers-Brillouin (WKB) technique is employed.

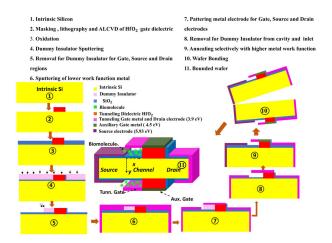


FIGURE 2. Proposal of fabrication flow diagram for dielectric modulated charge plasma based DM DGTFET.

The models and characteristics of the reported work [46] are also used to demonstrate the viability of the simulator in the suggested model. To validate the simulator, the drain characteristics are extracted from the reported paper using the plot digitizer tool. When the cavity is filled with air, the transfer characteristics of calibrated biosensor almost match the reported result, as exhibited in Fig. 3.

III. RESULTS AND DISCUSSION

When biomolecules are positioned into the cavity, the gate capacitance alters the interactions between the channel and the gate electrodes through electrostatic interaction. It perturbs various electrical characteristics like I_{ON}/I_{OFF} ratio,

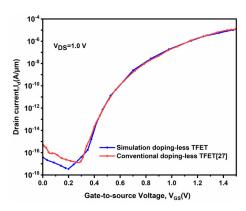


FIGURE 3. Drain current calibration of CP DM DGTFET biosensor compared with reported research work [46] at fixed $V_{DS}=0.1\ V_{CS}=0.1\ V_{CS}=0.1\$

subthreshold swing (SS), transfer characteristics, and energy band bending. In this section, minimum tunneling phenomena is noted for air filled cavity (k = 1) and is used throughout the literature to act as a constant reference value.

A. EFFECT OF k AND CHARGE OF BIOMOLECULES ON ENERGY BAND DIAGRAMS

The source - channel junction has a wide potential barrier width in the OFF state ($V_{DS}=0.5\ V$ and $V_{GS}=0\ V$) as noticed in Fig.4(a). The valence band of the source and the conduction band of the channel are not aligned in the OFF state (the dotted lines). Solid lines represent the energy band diagram in the ON state ($V_{DS} = 0.5 \text{ V}$ and $V_{GS} = +1.5 \text{ V}$). Due to the narrow tunneling barrier in this state, electrons engage in tunneling, and transition from the conduction band of the channel to the valence band of the source. The results are obtained for a positive charge biomolecule with dielectric constant k = 12 for a fixed cavity length of 20 nm. Fig. 4(b) shows a band diagram response for the ON and OFF state conditions for a positive and a negative charged biomolecule in the cavity with a constant relative permittivity k = 5 for a cavity length of 20 nm. The graph clearly shows that the positive charge biomolecule shifts the energy band downwards, while negative biomolecules shift the energy band upwards.

For the positive charge biomolecules, gate capacitance has more electrostatic control over intrinsic channel thus enhancing early commencement of tunneling phenomena. On the other hand, a fully filled cavity with negative charged biomolecules results in gentle misalignment of energy bands thus reducing tunneling phenomena at ON state and consequently drain current drops down.

B. EFFECT OF CAVITY LENGTH ON DRAIN CURRENT

The drain characteristics of CP DM DGTFET for the neutral biomolecule with a fixed relative permittivity k=6.3 for different cavity length are shown in Fig.5(a). It is noticed that for cavity length greater than 10 nm, the dependency of length of cavity on drain characteristics is not significantly pronounced. This is due to the presence of an effective work

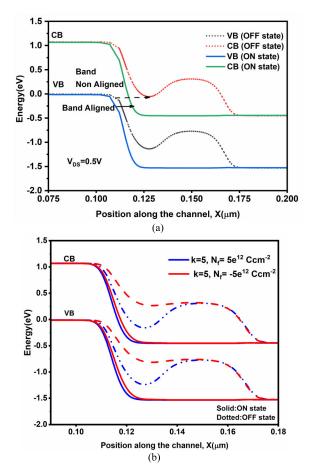


FIGURE 4. Band diagram for energy during the ON and OFF state conditions (a) Effect of the neutral biomolecule dielectric constant k=12. (b) Comparison of energy bands for biomolecules with negative and positive charges for the same dielectric constant k=5.

function metal over the junction area between the channel and the source. It is sufficient to trigger lateral tunneling. For reduced cavity length of 6 nm or less, the channel offers more resistance to tunneling current until it attains threshold voltage (which is 1.1 V). This is due to reduced covering area of lower work function metal. To prevail the tunneling phenomena, one requires larger area of lower metal work function of gate ϕ_t to increase the steepness at the tunneling junction. The I_{OFF} remains the same due to reduced ambipolarity behaviour owing to dual metal structure of gate. For higher cavity length $L_{cav}=25$ nm, the ambipolarity behaviour is found significantly less and so less band bending at channel-drain region eventually lower the I_{OFF} effectively. Hence the optimum choice of cavity length is taken as $L_{cav}=20$ nm for further investigation.

With fixed $L_{cav}=20$ nm, $V_{DS}=0.5$ V, drain characteristics with regard to a variety of biomolecules are plotted in Fig. 5(b) for the given structure. When a biomolecule of different dielectric constant (from Table 1) enters the cavity region, the energy barrier between the source and the channel narrows down, and hence I_{ON} rises. The biomolecules here act as a modulating parameter for controlling tunneling

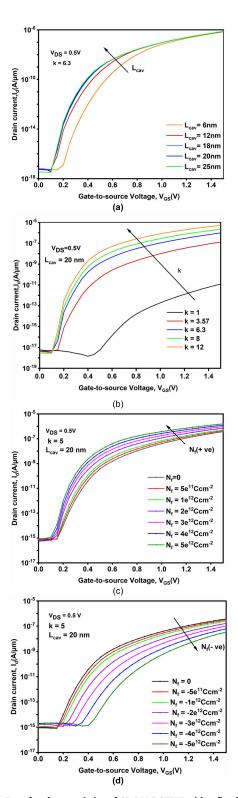


FIGURE 5. Transfer characteristics of CP DM DGTFET with a fixed V_{DS})=0.5 V (a) Cavity length, $L_{cav}=6$, 12, 18, 20 and 25 nm (b) Dielectric constant (k = 1, 3.57, 6.3, 8, and 12) (c) Positive charge distribution (ranging from 0 Ccm⁻² to 5×10^{-12} Ccm⁻²) for k = 5. (d) Negative charge distribution (ranging from 0 Ccm⁻² to -5×10^{-12} Ccm⁻²) for k = 5.

electrons from source to channel. As relative permittivity of neutral biomolecule increases, the electrostatic coupling of



gate voltage over lateral tunneling point increases, resulting I_{ON} to increase as shown in Fig. 5(b). The suggested structure can therefore identify biomolecules that have been calibrated against dielectric constant.

For positively charged biomolecules immobilized in the cavity with fixed $V_{DS}=0.5~V$ and fixed k=5, drain current variation is plotted in Fig. 5(c) for a fixed cavity length of $L_{cav}=20~\rm nm$. It has been noted that increased positive charge strengthens the tunneling phenomenon and hence raises the drain current. On the other hand, I_{OFF} remains unchanged as the channel-drain junction remains unaffected. For the similar reason, negative charged biomolecule reduces I_{ON} with increased negativity due to widening of tunneling barrier as depicted in Fig. 5(d).

C. EFFECT OF k ON DRAIN SENSITIVITY AND SELECTIVITY

Drain sensitivity is a crucial factor to detect various biomolecules. The drain sensitivity may be defined as the ratio of the drain current $I_{d(Bio)}$ when the cavity is completely occupied with biomolecules to the drain current $I_{d(air)}$ when it is vacant or occupied with air). Thus.

Drain Sensitivity =
$$\frac{Id(Bio)}{Id(air)}$$
 (2)

The drain sensitivity is studied for a fixed V_{DS} =0.5 V and with varying gate voltage, charge, cavity length and dielectric. The drain sensitivity is larger for positively charged biomolecules, as shown in Fig. 6(a). Higher positive charge molecules enhance the early onset of tunneling phenomena. The peak drain sensitivity increases from 6.5×10^7 to 1.3×10^9 as the charge distribution increases from neutral to 5×10^{12} Ccm⁻². Fig. 6(b) shows negative charge molecules reducing the tunneling phenomena by inducing gentle slope at the tunneling junction. Our studies reveal that the drain sensitivity shows a wide variation from 6.5×10^7 to 4.46×10^3 as the negativity of biomolecule changes from neutral to -5×10^{12} Ccm⁻². Hence, we conclude that the designed device is able to identify the presence and type of charge of biomolecules in a cavity. This is explained by the fact that the charges affect coupling of gate with the silicon channel and shifts energy band upside/ downside at the source- channel transition region for positive/ negative charges, respectively. In Fig. 6(c), we study drain sensitivity against various cavity lengths, and it shows that for longer cavity, the drain sensitivity is good enough to detect biomolecule while shorter cavity length results in shorter tunneling gate length hence lesser impact to enhance the ON current of the sensor. For cavity length 20 nm and 25 nm the results have been studied for neutral biomolecule with k = 6.3 and drain sensitivity is found to be almost the same. We discussed earlier that the energy band of the channel region drops down and the gate control effectiveness of the device becomes stronger for high-dielectric biomolecules. As evident from Fig. 6(d), for k = 12, the highest drain current sensitivity attained is 2.9×10^9 . Therefore, the tunneling probability function between the source and the channel is increased, and hence offers higher drain current, and consequently greater drain sensitivity as shown in Fig. 6(d).

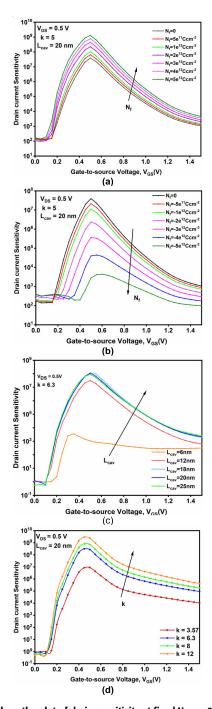


FIGURE 6. Show the plot of drain sensitivity at fixed $V_{DS}=0.5~V$ with variation of (a) Density of positive charges (which ranges from 0 Ccm-2 to $5\times 10^{-12}~Ccm^{-2}$ for fixed relative permittivity $k=5~and~L_{cav}=20~nm$. (b) Density of negative charges (which ranges from 0 Ccm⁻² to $-5\times 10^{-12}~Ccm^{-2}$ for fixed relative permittivity k=5, $L_{cav}=20~nm$ (c) Cavity length ($L_{cav}=6$, 12, 18, 20, 25 nm) for neutral biomolecule with k=6.3 (d) Relative permittivity of various neutral biomolecules (k=3.57, 6.3, 8, 12) for $L_{cav}=20~nm$.

Identifying a particular biomolecule from many other biomolecules is accomplished in terms of the parameter



"Selectivity" (ΔS) which is defined as under

$$\Delta S = \frac{I_d (k2) - I_d (k1)}{I_d (k1)}$$
 (3)

where k2, k1 are biomolecules of different dielectrics.

In our work, distinguishability of keratin (k = 8), from bacteriophage T7 (k = 6.3) can be obtained as

$$\Delta S_1 = \frac{I_d (k=8) - I_d (k=6.3)}{I_d (k=6.3)} = 1.27083.$$

Similarly, for the pair gelatin and keratin the selectivity is calculated as $\Delta S_2 = \frac{I_d(k=12) - I_d(k=8)}{I_d(k=8)} = 22.21$.

With the presented architecture, specific biomolecule binding happens through the cavity approach, which uses a label-free sensing technique based on dielectric modulation. This ensures that the amount of the targeted analyte is properly transduced to a proportionate modification of the output electric signals. A complete study of selectivity can be obtained by transient analysis of the device.

D. EFFECTS ON SUBTHRESHOLDSWING

The influence of neutral biomolecules on subthreshold swing (SS) is shown in the bar plot of Fig. 7 for given structure of cavity length $L_{cav}=20$ nm. When the relative permittivity of neutral biomolecules is varied from k=3.57 to 12, it shows variation from 28 mV/dec to 17 mV/dec, while air-filled cavity shows a SS of 70 mV/dec. The decrease in SS with dielectric constant is due to feeble effect of dielectric on tunneling junction for enhancing tunneling current, while dual metal work function enables the device to switch from ON to OFF state in a faster way. The minimum SS achieved for the device at k=12 is 17 mV/dec. As evident from Fig. 7, this device performs well at a low supply voltage $(V_{DS}=0.5\ V)$.

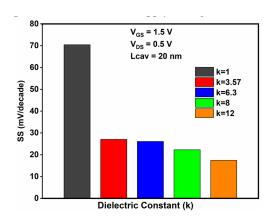


FIGURE 7. Subthreshold slope (SS) for various neutral biomolecules at $V_{GS}=1.5\ V,\ V_{DS}=0.5\ V$ and $L_{cav}=20\ nm.$

E. EFFECTS ON AVSS AND ION/IOFF

The average subthreshold slope(AVSS) is defined as

$$AVSS = \frac{V_t - V_{off}}{\log \left(I_d \left(V_t\right) - I_d \left(V_{off}\right)\right)}$$
(4)

where V_t is threshold voltage of the tunnel FET and V_{off} is the voltage V_{GS} when device is in OFF state. The drain current $I_d(V_{off})$ refers to the case when the device is OFF and $I_d(V_t)$ corresponds to the current at the threshold voltage. The effect of neutral biomolecules on AVSS have been studied and the result is plotted in Fig.8. (a). Similar to the explanation given in Fig. 6 (d), when dielectric constant (k) of neutral biomolecules increases, the I_{ON}/I_{OFF} ratio rises because I_{ON} increases without any change in the I_{OFF}. Also, it shows that when the dielectric constant gets higher, the average SS within the cavity falls. It is observed from Fig. 8 (a) that the I_{ON}/I_{OFF} ratio increases with higher k-values of the biomolecules due to the reasons explained earlier. Additionally, it is noted in Fig. 8 (a) that the AVSS decreases rapidly for k less than 4. A smaller SS obtained from this biosensor dictates a better electrical response and an easy ability of detection.

For charged biomolecules, the I_{ON}/I_{OFF} ratio and average subthreshold slope (AVSS) is plotted in Fig. 8(b) with respect to the variation of charges from negative to positive. At tunneling junction, as the positive value of charge increases, the I_{ON} current is enhanced due to steep band bending of energy bands. Consequently, the current ratio and AVSS increases.

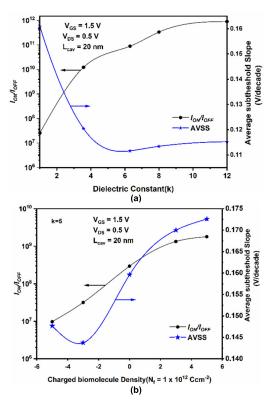


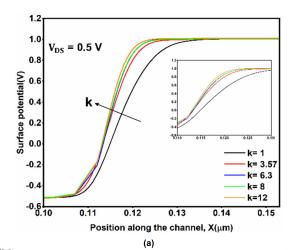
FIGURE 8. Variation of AVSS, and I_{ON}/I_{OFF} ratio of Charge Plasma based DM DGTFET for (a) Neutral biomolecules for $L_{cav}=20$ nm (b) Charged molecules of constant permittivity k=5 and $L_{cav}=20$ nm.

F. EFFECT OF k ON SURFACE POTENTIAL

Investigating the surface potential beneath the nanogap cavity is vital to grasping the electrostatic potential behaviour of



the charge plasma-based DM DGTFET biosensor. The surface potential along the channel corresponding to different dielectric constants is illustrated in Fig. 9(a). It is shown that when biomolecules are absent (k = 1), the surface potential is lowest. The reason for this is that a higher dielectric constant causes a stronger coupling between the gate and the channel, which in turn causes the barrier width at the source–channel interface to decrease. This effectively raises the surface potential, for k > 1. Examining the nano system channel region, one observes contour waves of electrostatic potential based on the movement of carriers which begins at the source and ends at the drain as shown in Fig. 9(b). The presence of neutral biomolecules creates strong potential contours in the cavity region, thereby influencing the tunneling of holes in the source-channel interface. This is due to the additional influence of dielectric of the biomolecules.



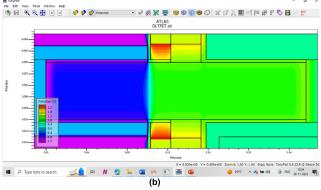


FIGURE 9. (a) Surface potential along the channel for various biomolecules at constant $V_{DS}=0.5$ V. (b) Electrostatic potential contours observed across the channel at constant $V_{DS}=0.5$ V, $V_{GS}=1.5$ V, and for k=12.

G. EFFECTS OF k AND CHARGES ON DEVICE EFFICIENCY

Another pertinent parameter that can measure the device's capability to convert drain current I_d into g_m , is the device efficiency, $\left|\frac{g_m}{I_d}\right|$. Here, gm denotes the transconductance of a device and is given as

$$g_{m} = \left. \frac{dI_{d}}{dV_{GS}} \right|_{V_{DS=Constant}} \eqno(5)$$

A rise in the maximum value of $|g_m/I_d|_{max}$ is the feature that most clearly distinguishes distinct biomolecules. The spatial shifts in subthreshold swing and $|g_m/I_d|_{max}$ is of different magnitudes for different biomolecules.

The effect of device efficiency $|g_m|/I_d$ as a sensing metric for the suggested biosensor is depicted in Figures. 10(a-e). It is established that as biomolecules build up, capacitance between the gate and the channel rises. Hence it improves the electric field, which may then be used to regulate the energy bands to enable carrier tunneling and drain current. It is evident in Fig. 10 (a-d), that at lower drain current, the transconductance to the current ratio (g_m/I_d) is comparatively higher because the presence of biomolecules eventually overcoming the tunneling barrier. After attaining higher gate voltage (V_{GS} < 1 V), as the transconductance decreases, drain current saturates, hence negative differential resistance (NDR) and negative differential transconductance (NDT) are observed in Fig. 10 (a). The quantum mechanical phenomena, NDT reflects the change of g_m from positive to negative values. In the absence of biomolecules (i.e. filled with air) the onset of tunneling phenomena could not be possible hence the controlling of drain current reduces. Gate architecture is maintained with fixed EOT throughout, but the presence of biomolecules at source-channel interface modulates NDT, thus enabling to recognize different biomolecules. Positively charged molecules, for the same reason, show a higher $|g_m/I_d|_{max}$ at the lower drain current Fig. 10(b). It would be easier to detect different biomolecules by knowing the shift in the maximum device efficiency at constant current values. Similarly, for negative biomolecules, the device efficiency decreases almost proportionately, as shown in Fig. 10(c).

Fig. 10(d) shows the variation of device efficiency with drain current for different cavity lengths. Higher cavity lengths, increases the tunneling gate length. Thus, with low drain current, the electric field at the source - channel interface promotes the start of tunneling. An optimized device structure with neutral biomolecules (k = 6.3) is thus found with a cavity length of 20 nm. For higher dielectric biomolecules, one can observe the tunneling phenomena predominates at lower drain current and operating V_{GS} as $|gm/Id|_{max}$ shifts leftwards as shown in Fig.10(e). In Fig. 10(e), the plot of device efficiency against V_{GS} is shown. It emphasizes that neutral molecules with $k=3.57,\,6.3,\,8,$ and 12, achieve $|g_m/I_d|_{max}$ at V_{GS} equals 0.15 V approximately. At lower voltages, the tunneling switches from OFF to ON state.

H. EFFECTS OF k ON ELECTRIC FIELD AND BAND TO BAND TUNELING (BTBT) RATE

Figure 11 illustrates the electric field profile for various neutral biomolecules. It has been noted that when biomolecules' dielectric properties increase, so does the electric field. This occurs when the suggested device gate voltage is 1.5~V for k=12, as discussed in Fig. 10(e). When there is no biomolecule present in the cavity, an electric field of around 1.17~MV/cm is produced; when gelatin is present, a maximum electric field of around 1.96~MV/cm grows up across

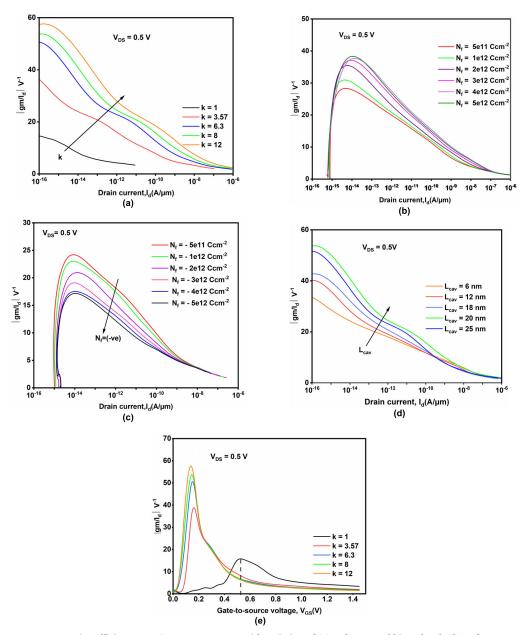


FIGURE 10. Device efficiency $|g_m/I_d|$ at $V_{DS}=0.5$ V with variation of (a) I_d for neutral biomolecule (b) I_d for positive charge biomolecule (c) I_d for negative charge biomolecule (d) I_d for cavity length variation (e) V_{GS} for different neutral biomolecule.

the channel. The higher the value of k, the narrower is the bell-shaped response just above the cavity due to additional electrostatic control at the junction.

The band to band tunneling rate (BTBT) for different biomolecules is shown in Fig. 12. The electron tunneling rate rises with the dielectric values. This behaviour can be explained in the light of high capacitive coupling for higher k-values that lead to more energy band alignment favoring high tunneling.

I. RADIO-FREQUENCY ANALYSIS

The evaluation of the radio frequency (RF) performances of the given structure for various biomolecules is also provided, as it is vital for maintaining low dynamic power consumption. The characteristics of gain bandwidth product (GBP), transconductance (g_m) , gate-to-drain capacitance (C_{gd}) , and cut-off frequency (f_T) are measured in this work. The capability to convert the given gate voltage into drain current is defined by the parameter transconductance (g_m) .

Gate-to-drain capacitance (C_{gd}) is a parasitic capacitance in the devices, crucial for radio-frequency applications. Analyzing C_{gd} is essential to study switching behaviour and power dissipation of the device in circuits. It is defined by

$$C_{gd} = \frac{\partial Q_g}{\partial V_{DS}} \tag{6}$$

where Q_g is the charge on gate.



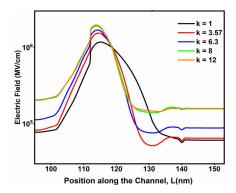


FIGURE 11. Variation of electric field observed across the channel at constant $V_{DS}=0.5\ V,\ V_{GS}=1.5\ V.$

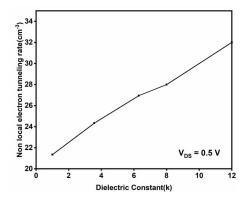


FIGURE 12. Electron tunneling rate for different biomolecules at constant VDS = 0.5 V, and VGS = 1.5 V.

Figure 13(a) shows that transconductance with the neutral biomolecules increases with the dielectric constants; g_m shows almost linear dependence with gate voltage for all k- values for, $V_{GS}>0.7$ V. The variation of C_{gd} with gate voltage for various biomolecules has been plotted in Fig. 13(b) and indicates that an increase in dielectric results in a reduction of capacitive coupling at channel-drain region. This is due to the lower density of states, making it favorable for high-frequency regime. The charge builds up at the drain junction; hence, the total parasitic capacitance increases with gate voltage.

At a voltage close to 0.7~V, there is a rapid increase in g_m . The cut-off frequency (f_T) , which denotes the device can act as a unity-gain current amplifier is a further aspect for quicker switching applications. It is given by

$$f_{\rm T} = \frac{g_{\rm m}}{2\pi C_{\rm gg}} \tag{7}$$

where $C_{gg} = C_{gs} + C_{gd}$, and C_{gs} is the depletion capacitances source with respect to gate. Fig. 13(c) demonstrates the variation of f_T for neutral biomolecules in the cavity. Induction of tunneling at source – channel region due to the high-k dielectric materials results in higher cut off frequency. Near linear increase in cut off frequency with the gate voltage (>0.7V) can be explained in the light of nature

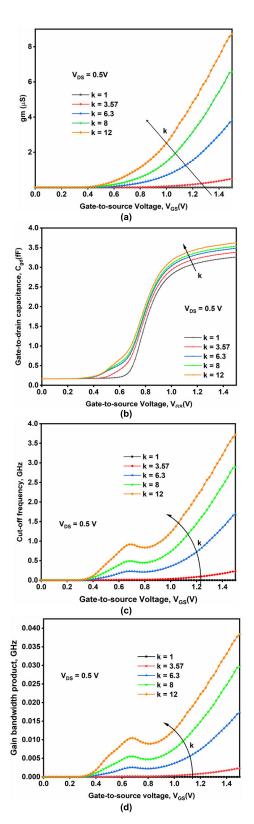


FIGURE 13. RF analysis with constant V $_{DS}=0.5$ V (a) g_m (b) C_{gd} (c) f_T (d) GBP for various neutral biomolecule.

of variation of C_{gd} of Fig. 13(b). As g_m continues increasing more steeply for CPTFET, and parasitic capacitance



builds up at the channel- drain region till $V_{GS}=1~V$, the f_T going on increasing linearly as expected. In absence of biomolecule (k=1), there is large tunnel gap at source junction, hence feeble possibility of tunneling of hole. Thus, increase in cut off frequency to be utilized for detection of biomolecules.

The gain bandwidth product has similar behaviour as f_T as plotted in Fig. 13(d). It is the product of bandwidth and gain, and it represents the device's working region over its steady gain region. It is used in the high frequency region and is expressed as

$$GBP = \frac{g_{\rm m}}{20\pi C_{\rm od}} \tag{8}$$

Another metric for studying high-speed operation is the transit time (t_t) . The duration required for charge carriers to traverse the channel region is referred to as the transit time (t_t) and can be expressed as follows:

$$t_t = \frac{1}{2\pi f_T} \tag{9}$$

A shorter transit time leads to faster operation of the transistor, contributing to improved switching speeds and overall device performance. The transit times (t_t) for distinct neutral biomolecules are shown in Figure 14. The device shows higher sensing speed for high k dielectric biomolecules like gelatin. Charges swipe through the channel; hence higher dielectric induces narrowing of tunneling barrier with increase in the gate voltage.

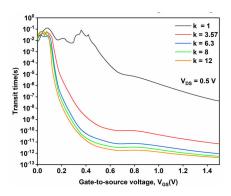


FIGURE 14. Variation of t_t with $\rm V_{GS}$ for various neutral biomolecules at constant $\rm V_{DS}=0.5~\rm V.$

J. LINEARITY ANALYSIS

Minimal signal distortion is crucial for maintaining the integrity and quality of the signal as far as the high-frequency transport of carriers is concerned. Hence linearity analysis has been carried to investigate its performance while working in radio frequency regime. The device linearity is studied here for various biomolecules in term of the parameters such as voltage intercept point(VIP), Input intercept point(IIP), and Intermodulation Distortion(IMD), as defined

below:

$$g_{mn} = \frac{1}{n!} \frac{\delta^n I_d}{\delta V_{GS}^n}$$
 (10)

$$VIP2 = 4 \left(\frac{g_{m1}}{g_{m2}} \right) \tag{11}$$

$$VIP3 = \sqrt{24 \times \left(\frac{g_{m1}}{g_{m3}}\right)} \tag{12}$$

$$IIP3 = \frac{2}{3} \times \left(\frac{g_{m1}}{g_{m3} \times R_s}\right) \tag{13}$$

$$IMD3 = \left[\frac{9}{2} \times (VIP3)^2 \times g_{m3}\right]^2 \times R_s$$
 (14)

where transconductance is represented as gm, and 'n' denotes the order which can be n=1, 2, 3... and the value of R_s is usually taken as 50 Ω for RF application. Second-order distortion (proportional to g_{m2}) generates harmonics and intermodulation products at twice the input frequency, causing cross-modulation. Third-order distortion (proportional to g_{m3}) creates in-band intermodulation products, leading to significant interference within the system's bandwidth. For any device, higher-order parameters such as VIP2, VIP3 and IIP3 should be large as possible for achieving linearity with respect to various DC parameters. The plot of VIP2, VIP3 and IIP3 with respect gate to source voltage for various biomolecules at fixed bias of $V_{DS} = 0.5V$ are shown in Figure 15(a), (b) and (c), respectively. Biomolecules with higher k shows more linearity metrics, as depicted in Fig. 15(a). With a high k biomolecule, the tunneling is more and g_m attains a constant value thus providing more linear response. The metric VIP3 is more important as it involves gm of third order. VIP3 for various biomolecules, with k=1, 3.57, 6.3, 8 and 12 are plotted in fig 15(b). VIP3 with airfilled cavity(k=1) is shown in subplot (i) of Fig.15(b) and so on. The variation of VIP3 with respect to the applied gate bias shows peaks occurring at different bias voltage for different biomolecules. When the cavity is filled with high-k biomolecule, the device shows less distortion. Another way to asses this nonlinearity is the measure of IIP3 as shown in Fig. 15(c). It is obvious that the peaks are observed beyond saturation for all the studied biomolecules.

The variation of IMD3 with input gate voltage is shown in figure 15(d). It affirms that the device is poorly effected from non-linearity while detecting biomolecules. Lower the value of IMD3, the less is the effect of nonlinearity.

The drain current values obtained by the present devices for different biomolecules are compared with the previously reported biosensor works [1], [7], [14], [15], [38] in Table 3. The values of the published works are acquired and are compared with the neutral biomolecule present in the cavity of the present devices. The biosensor exhibits enriched sensitivity than other charged biomolecules at lower operating voltages instigating the fact that the present biosensor devices designed here are much healthier providing enhanced performances.



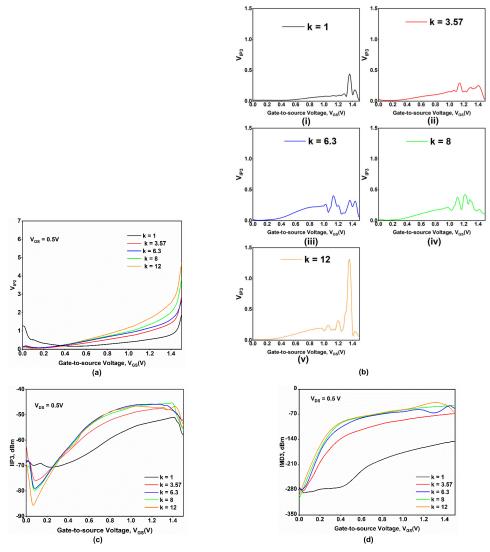


FIGURE 15. Variation of (a) VIP2, (b)VIP3, (c) IIP3 and (d) IMD3 with constant $V_{DS} = 0.5 \text{ V}$ for various neutral biomolecules.

TABLE 3. Comparison of fet-based biosensor performance with previously reported works.

	Targeted Biomolecules (K values)	Present work	Reported works
Drain Current	Neutral 3.57	1.00 ×10 ⁻⁶	5.11562×10 ⁻⁹ [37]
(A/μm) at	Neutral 6.3	5.13×10^{-6}	2.01×10 ⁻⁷ [37]
$(V_{GS} = 1.5 \text{ V})$	Neutral 8	1.17×10^{-5}	9.17×10 ⁻⁷ [37]
& $(V_{DS} =$	Neutral 12	2.43 × 10 ⁻⁵	5.31801×10 ⁻⁶ [37]
1 V)	Neutral 12	2.43 × 10 °	4.00374×10 ⁻⁶ [1]
I_{ON}/I_{OFF} ratio at			8.09×10 ¹¹ [37]
$(V_{GS}: 1.5 \text{ V}) \&$	Neutral 12	9.02×10^{11}	1.45×108 [14]
$(V_{DS} : 1 V)$			3.77 ×10 ⁹ [15]
Subthreshold Swing (SS)	Neutral 12	17mv/dec	21.7 mV/dec [37]
Drain current (A/µm) at	k = 6.3	4.99 × 10 ⁻⁶	$3.9 \times 10^8 [15]$
(V _{GS} : 1.2 V) & (V _{DS} : 1 V)	k = 12	2.13 × 10 ⁻⁵	3.76 × 10 ⁻⁸ [15]
Drain current	k = 8	3.54×10^{-6}	2.20 x 10 ⁻⁷ [7]
(A/µm) at (V _{GS} : 1.2 V) & (V _{DS} : 0.5 V)	k = 12	8.02 × 10 ⁻⁶	9 x 10 ⁻⁷ [7]

IV. CONCLUSION

A charge plasma-based DM DGTFET for the detection of label-free biomolecules has been explored and analyzed using dielectric modulation in the cavity beneath the tunnel gate metal. Electrostatic features like energy band diagram, drain characteristics, drain current sensitivity, subthreshold swing, and RF performance have been investigated for the biosensor. The energy band diagram in the ON and OFF state conditions for the device is explored for both neutral, positive, and negative charge biomolecules. A detailed investigation reveals that lower cavity lengths (~ 6 nm) require a higher gate voltage to achieve drain current than higher cavity lengths (> 12 nm). Drain current saturates at higher $V_{\rm GS}$ for all cavity dimensions, and with the optimal cavity length at 20 nm, drain sensitivity is at its maximum. The highest drain current sensitivity of 1.3×10^9 is attained for positive charge biomolecules



and 2.9×10^9 for neutral biomolecules (k = 12). Notably, the peak of the drain sensitivity shows a wide variation with respect to the charges of the biomolecules. Also the cavity length has profound impact on the drain current sensitivity. The study shows that the subthreshold swing (SS) decreases for higher k, showing a maximum of 70 mV/dec for an airfilled (k = 1) cavity sensor. The I_{ON}/I_{OFF} ratio increases with both k and charges of biomolecules since I_{ON} increases without significant change in I_{OFF} . Here, a proper calibration has led to the detection of the charge on the biomolecule over the range between -5×10^{12} to 5×10^{12} Ccm⁻². AVSS also decreases negative charge densities of biomolecules. So, a variation of AVSS with k suggests easy detection of biomolecules in the sensor.

The channel electric field profile is bell-shaped as usual; a high k narrows down the profile, establishing strong electrostatic control on the charge transport in the channel. Also, the band-to-band tunneling rate is higher for higher k. The sensor's RF performance is evaluated, and it is found that both gm and C_{gd} increase with $V_{GS}(> 0.4 \text{ V})$ and dielectric constant k, as expected. The cut-off frequency f_T increases almost linearly with the gate voltage (> 0.7 V), as does the gain bandwidth product. To distinguish biomolecules from others, metric selectivity is defined and discussed. Finally, the linearity analysis for the device is investigated by studying the higher orders of VIP, IIP, and IMD. The device becomes more linear and stable for biomolecules with high k values. The RF analysis covers the transit time, cutoff frequency, and gain bandwidth product and shows that the device is faster for detecting neutral and charged biomolecules. With robust electrostatic control, enhanced tunneling rates, and favorable RF performance metrics, the split-dielectric CP DM DGTFET emerges as a promising solution for biosensing applications.

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